

The adaptive radiation of lichen-forming Teloschistaceae is associated with sunscreens pigments and a bark-to-rock substrate shift

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Edited by David M. Hillis, The University of Texas at Austin, TX, and approved July 15, 2015 (received for review April 10, 2015)

Adaptive radiations play key roles in the generation of biodiversity and biological novelty, and therefore understanding the factors that drive them remains one of the most important challenges of evolutionary biology. Although both intrinsic innovations and extrinsic ecological opportunities contribute to diversification bursts, few studies have looked at the synergistic effect of such factors. Here we investigate the Teloschistales (Ascomycota), a group of >1,000 lichenized species with variation in species richness and phenotypic traits that hinted at a potential adaptive radiation. We found evidence for a dramatic increase in diversification rate for one of four families within this order—Teloschistaceae—which occurred ~100 Mya (Late Cretaceous) and was associated with a switch from bark to rock and from shady to sun-exposed habitats. This adaptation to sunny habitats is likely to have been enabled by a contemporaneous key novel phenotypic innovation: the production in both vegetative structure (thallus) and fruiting body (apothecia) of anthraquinones, secondary metabolites known to protect against UV light. We found that the two ecological factors (sun exposure and rock substrate) and the phenotypic innovation (anthraquinones in the thallus) were all significant when testing for state-dependent shifts in diversification rates, and together they seem likely to be responsible for the success of the Teloschistaceae, one of the largest lichen-forming fungal lineages. Our results support the idea that adaptive radiations are driven not by a single factor or key innovation, but require a serendipitous combination of both intrinsic biotic and extrinsic abiotic and ecological factors.

adaptive radiation | lichens | sunlight protection | substrate switch | Teloschistaceae

The rate at which species proliferate through evolutionary time varies greatly from one evolutionary lineage to another, forming one of the most notable characteristics of macroevolutionary change. Adaptive radiations, in which a period of increased speciation is caused by adaptation to novel environments, are particularly important in generating biological diversity (1–3). Understanding the drivers behind these events is thus an important goal. Two basic categories of drivers have been described: extrinsic ecological opportunities that are presented to the ancestors of radiations (e.g., refs. 4 and 5) and intrinsic evolutionary adaptations within the ancestral lineage that result in breakthroughs that spur diversification (6–10). At an extreme, the latter may involve only one trait—a “key innovation” (11–13). Although both processes are likely to be involved in any adaptive radiation, there have been few attempts to investigate how they may be entwined (14, 15): innovations may immediately generate an ecological opportunity by providing access to a new ecological niche; conversely, the exploitation of a new opportunity might generate strong directional selection, leading to rapid evolutionary change at the origin of the adaptation, the results of which permeate throughout the entire lineage.

A potential example of an evolutionary radiation can be found in a group of lichens [obligate mutualistic ectosymbioses between fungi and either or both green algae and cyanobacteria (photobionts) (16)]: the family Teloschistaceae. Although cosmopolitan, this taxon is especially associated with exposed habitats, being able to colonize some of the most arid regions of the world. Because many lichens can use fog and dew as their principal source of water and can be highly tolerant to desiccation, they might be regarded as well adapted to arid exposed habitats. However, many groups of lichens are limited in their ability to exploit such habitats, due, in part, to the high levels of UV-light exposure. Protection against excessive irradiation is required (17), especially for the photobiont partner (18, 19).

In the Teloschistaceae, the solution to this problem lies in their characteristic orange and yellow pigmentation, which results from the accumulation of cortical anthraquinones. These secondary metabolites are produced by the fungal partner, and their crystals alter the spectral composition of the light that goes through the cortex (20) by absorbing visible blue light, UV-B and -A radiations (21, 22). Synthesis of anthraquinones and other lichen pigments is stimulated by UV-B radiation (e.g., refs. 23–26), and they are thought to reduce UV-B-induced damage to

Significance

The tempo of diversification of life can be accelerated by fortuitous ecological opportunity or by phenotypic innovation. In this study, we document how both factors are likely to have played a role in the origin and success of a major fungal lineage, the Teloschistaceae (comprising ~1% of all fungi). Anthraquinone pigments are found in a widespread, but scattered, range of fungi and plants, but are particularly abundant in the lichen-forming Teloschistaceae, where they provide sunlight protection, especially needed when growing in arid deserts of the world. We found that anthraquinones evolved in these lichens, in conjunction with an ecological switch to exposed, rocky environments, allowing them to colonize swathes of unexploited habitats worldwide and sparking an acceleration in diversification.

Author contributions: E.G. and F.L. designed research; E.G., S.F.-B., R.V., R.F.L., M.R.-M., and F.L. performed research; E.G., R.F.L., and F.L. contributed new reagents/analytic tools; E.G., S.F.-B., R.F.L., and C.G. analyzed data; and E.G., R.F.L., C.G., and F.L. wrote the paper. The authors declare no conflict of interest.

This article is a PNAS Direct Submission.

Data deposition: The sequences reported in this paper have been deposited in the GenBank database (accession nos. [KT291442–KT291669](https://doi.org/10.1073/pnas.1507072112)). Sequence alignment and tree file data are available from TreeBASE, treebase.org (Study id no. 18015).

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This article contains supporting information online at www.pnas.org/lookup/suppl/doi:10.1073/pnas.1507072112/-DCSupplemental.

DNA in both the fungus and the alga of lichens (22). Anthraquinones are polyketide aromatic compounds, derived from the acetyl polymalonyl pathway and synthesized by multidomain polyketide synthases (PKSs), which are encoded by PKS gene clusters (19). Only one gene involved in anthraquinone synthesis in lichens has been characterized so far: the transcript of a nonreducing fungal type I PKS gene from the cultured mycobiont of *Xanthoria elegans* (27). There are various closely related anthraquinones in the Teloschistales, and each species may possess several in different proportions. The pattern of variation among taxa is still poorly understood, appears to have little phylogenetic signal, and may depend on environmental as well as genetic factors. Because all appear to serve the same function, here we simply consider the presence or absence of anthraquinones. The ubiquity of anthraquinones within the Teloschistaceae

is, however, unique and hints at some evolutionary success favoring the production of these metabolites.

The Teloschistaceae, with ~650 described species (28) and an estimate of >1,000 extant species (29), shares common ancestry with three much less speciose families within the order Teloschistales: Brigantiaeaceae, Letrouitiaceae, and Megalosporaceae [<100 described species combined (30)]. Whereas the Teloschistaceae is a mainly rock-dwelling cosmopolitan clade with a diversity of growth habits ranging from crustose to fruticose, the Brigantiaeaceae and Letrouitiaceae (with a circumpacific center of diversity), as well as the Megalosporaceae (with an Australasian center of diversity), are generally epiphytic and crustose and are found mostly in the tropics (30).

In this study, we test the hypothesis that the high species richness of the Teloschistaceae is the result of an adaptive radiation, by

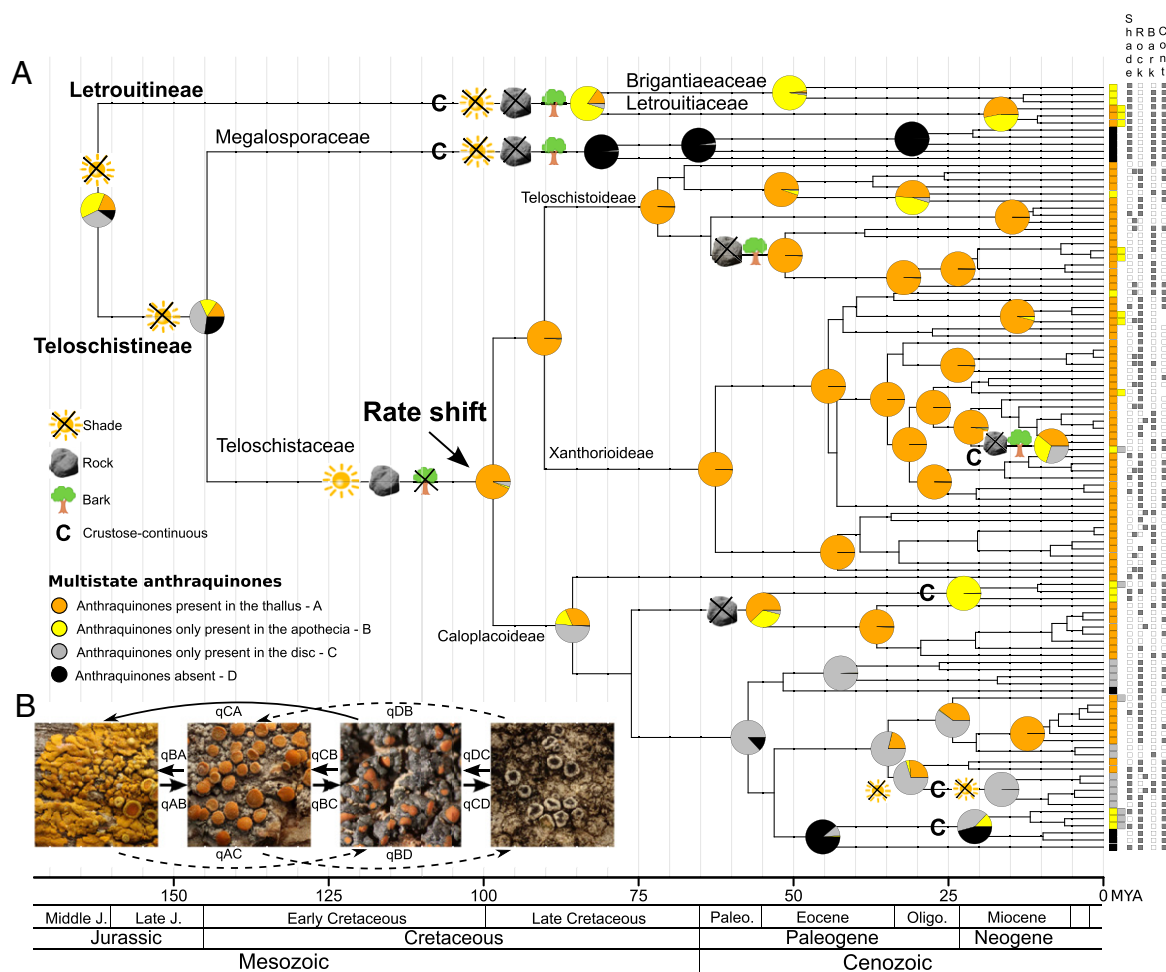


Fig. 1. Time-calibrated phylogeny of the order Teloschistales depicting multistate ancestral character state reconstructions of anthraquinone evolution. (A) Pies represent probabilities of each ancestor being in each of the four potential states for the presence/absence of anthraquinones. Colored squares at the tips of the tree indicate anthraquinone character states for extant taxa using the same color scheme as for the pie charts. Polymorphic taxa have more than one colored square. In addition, we mapped the presence/absence of other characters with symbols for ancestral states (on branches) and with black and white boxes for extant taxa (at the tips of the tree): a sun for light exposure (shade column), a rock for rock substrate (rock column), a tree for epiphytism (bark column), and a letter C for growth form crustose-continuous (cont column). These same symbols when crossed represent absence of the trait. We only show these symbols on the topology itself if we established high confidence in the state of the trait at a node and if we inferred a gain or loss of the trait at that particular node. These characters were coded and analyzed as binary character states (SI Appendix, Fig. S4). Geological times are indicated at the axis of the tree. This reconstruction illustrates a full anthraquinone ancestor living in sunny and rocky habitats as the most probable state at the crown node of the large family Teloschistaceae. These features are maintained throughout much of the history of the family. The same internode was significantly characterized by a major increase in diversification rates (Fig. 2). (B) Evolutionary model selected for the anthraquinone multistate reconstruction. Photographs illustrate the four different states of anthraquinone presence/absence in the lichen. Arrows indicate transitions between the possible character states. In this model, transitions and reversals are treated as equal, but single-step transitions and qCA (solid arrows) have higher rates than the remaining two-step transitions (dashed arrows), and three-step transitions are not allowed.

valuable portion of the apothecium (which contains the ascospores), until anthraquinones disappeared completely. Only one clade in the Caloplacoideae (*Caloplaca aurantia* group; *SI Appendix, Fig. S4A*) seems to have experienced a regain of anthraquinones in the thallus from a state of only disk presence.

Although our analyses show that the spread of anthraquinones throughout the thallus originated at the split of the family Teloschistaceae, the original acquisition of anthraquinones per se most likely took place at the root of the Teloschistales, as shown in *SI Appendix, Fig. S4B*, and was secondarily lost in the Megalosporaceae. This conclusion is reinforced by the fact that these pigments are mostly absent from the closest relatives to the Teloschistales (i.e., Caliciales, Lecanorales). The uncertainty in the multistate analysis (Fig. 1) might be an artifact of how we coded a continuously varying trait: whereas most Letrouitineae possess anthraquinones only in their apothecia, a few species of *Letrouitia* also have anthraquinones in their thalli, but compared with the Teloschistaceae, these pigments are far less abundant and often disappear completely in shady habitats.

Exposure to shade was estimated as the ancestral state at the root of the Teloschistales and has been maintained throughout the tropical Letrouitineae and also in the Megalosporaceae. This trait was lost—corresponding to a shift from shaded to sunlit habitats—along the stem lineage of the Teloschistaceae (Fig. 1 and *SI Appendix, Fig. S4*). Subsequently, there were several secondary gains, mostly within the Caloplacoideae, that were not recovered by all reconstruction methods.

Following the same pattern, the colonization of rocky substrates evolved during the early history of the Teloschistaceae and spread throughout the family, although it was secondarily lost on a few occasions (i.e., *Fulgensia* and *Teloschistes* clades; Fig. 1 and *SI Appendix, Fig. S4D*). The reconstruction of bark (epiphytism) as a substrate preference complements that of rock colonization. Although there is considerable uncertainty at the root, the predominance of bark as a substrate for tropical Letrouitineae and Megalosporaceae and its loss associated with the split of the Teloschistaceae is unequivocal. Most clades that showed secondary losses of rock as a substrate were replaced by secondary gains of bark, with the exception of the *Fulgensia* clade, where species mostly grow on soil and mosses (Fig. 1 and *SI Appendix, Fig. S4E*).

The evolution of growth forms (e.g., crustose, foliose, and fruticose thalli) showed the most discrepancies among methods and least well-supported ancestral estimation. We found that the trait crustose-continuous thallus seems to have been present at the origin of the tropical families (Fig. 1): although BayesTraits and *corHMM* approaches support the hypothesis that the trait was ancestral to the Teloschistales as a whole, lost in certain clades within the Teloschistaceae, and secondarily gained in a few instances, our BiSSE analysis supports several independent gains within the family (*SI Appendix, Fig. S4*). The reconstruction of attachment structures, which is associated with the evolution of the growth forms foliose and fruticose (i.e., a greater level of separation of the thallus from its substrate, compared with crustose lichens), was also quite uncertain. It seems plausible that the trait evolved repeatedly in several clades within the Teloschistaceae.

In summary, the origin of the Teloschistaceae corresponds with interrelated transitions in the phenotypic evolution of anthraquinones spreading through the thallus and the ecological evolution from shaded epiphytic habitats to exposed rocky substrates.

Testing for State-Dependent Diversification Rates. The origin of the Teloschistaceae was associated with both an increase in the rate of diversification and the evolution of key phenotypic traits. We therefore investigated more directly whether there was a link between these phenotypic traits and the shift in diversification rates, using BiSSE and accounting for the incomplete sampling of taxa in our dataset (41).

We found evidence that presence of anthraquinones in the thallus increased the rate of diversification, whereas living in shady habitats or possessing a crustose-continuous growth form both decreased the rate of diversification. In the first case, the effect was significant only when we assumed that 70% or fewer of species had been sampled ($\Delta\text{AIC} = 2.2$; $P = 0.04$ for 70% sampling), which is the most reasonable assumption: current estimates are of at least 1,000 species (29). In the case of shade and crustose-continuous thalli, the effect was significant even when we assumed all species were sampled ($\Delta\text{AIC} = 7$, $P = 0.003$ for shade; $\Delta\text{AIC} = 12$, $P = 0.0003$ in crustose-continuous for 100% sampling). In contrast, we found no evidence that substrate (rock or bark) or attachment structures influenced the rate of diversification (*SI Appendix, Table S1*).

Evolutionary shifts in these phenotypic traits may also be directly related to cladogenesis [sensu (42)], where character changes are associated with speciation events. We tested for cladogenetic effects using BiSSEness (42). Here we found evidence for significant cladogenetic effects with shade and with the rock and bark substrates (*SI Appendix, Table S2*), but not with anthraquinones nor with crustose-continuous growth forms or with attachment structures.

In summary, our results support the hypothesis that phenotypic innovations allow diversification through the exploitation of novel habitats. The presence of anthraquinones in the thallus and the colonization of rocks in unshaded habitats have both been central to the success of the Teloschistaceae. Beyond that, our results also indicate that phenotypic traits (anthraquinones and crustose-continuous) are especially correlated with diversification rates, whereas ecological substrate traits (rock and bark) are particularly linked to cladogenesis.

Setting Events in a Time Frame. Our results demonstrate an adaptive radiation, beginning at the onset of the diversification of the large Teloschistaceae family, and associated with a shift to sunny, rocky habitats and the innovation of using anthraquinones as a sun-protecting pigment. These events are likely to have occurred in the Early Cretaceous, ~100 Mya, during the Cretaceous Thermal Maximum (112–89 Mya). Several events seem to concentrate around that time. The onset of diversification of the Teloschistaceae could be linked to the increase in temperatures in the Cretaceous, leading to the colonization of newly formed arid and exposed regions, possibly augmented by greater provincialization (43) and changes in orographic landmarks, such as the separation of America and Africa (119–105 Mya) (44). Alternatively, the greater seasonality and the spread of deciduousness in both angiosperms and gymnosperms in the cooling period that followed the Cretaceous Thermal Maximum (43) may have changed the light environment for the epiphytic ancestors of the Teloschistaceae, leading to adaptation to higher light exposure. In this latter scenario, the intermediate light exposure offered by deciduous habitats might be seen as providing a “stepping stone” in the adaptive landscape, leading to the evolution of light-protective anthraquinones throughout the thallus and allowing the subsequent colonization of fully exposed habitats.

Several other examples of diversification events have been found in epiphytic groups, during the same period as the diversification of the Teloschistales, subsequent to the radiation of angiosperms [194 (210–162) Mya (45), 217 (257–182) Mya (46, 47), and 240–175 Mya (48)], suggesting that the terrestrial dominance of the latter in the Cretaceous, and the advent of the Pentapetalae radiation ~110 (115–105) Mya (45), triggered an ecological opportunistic response, including some fern groups (49, 50), epyphitic Lycophytes (51), and other groups of lichenized fungi [Lecanoromycetidae and Ostropomycetidae (52)]. However, in all of the above-mentioned studies, diversification is linked to the exploitation of novel epiphytic niches generated by angiosperms' more diverse and structured habitats and by the expansion of rain forests. The radiation of

the Teloschistaceae stands out in suggesting a contemporaneous move away from epiphytic habitats to saxicolous (rock-dwelling) ones, which leaves us with several open questions. Were the nascent Teloschistaceae outcompeted in the newly arisen forest canopies and forced to occupy more extreme but available habitats? In this scenario, anthraquinones evolved to occur in the thallus as a relatively trivial consequence of the exposed conditions. Or did the innovation of anthraquinones being present in the thallus allow the opportunistic colonization of exposed rocky habitats and accelerate diversification? In line with the first of these hypotheses, the sister groups of the Teloschistaceae (Letroitiaceae and Megalosporaceae) do not show signs of diversification outbursts and remain species-poor, despite being epiphytic, and predominantly tropical in distribution. Conversely, the facts that anthraquinones are found in only a very few other lichen species and that other successful lichen lineages were adopting epiphytism at the same time as the Teloschistaceae were colonizing open habitats point to the idea that anthraquinones were important factors in the taxon's evolutionary radiation.

These considerations—along with our direct findings that anthraquinones and ecological traits share a similar evolutionary history and are both related to diversification—lead us to conclude that rather than one or other playing a dominant role, ecological opportunities and phenotypic innovations are intricately entwined as factors causing adaptive radiations. On the one hand, anthraquinones represent a clear case of an innovation—a phenotypic trait that lies at the heart of the radiation. On the other hand, it is equally clear that this innovation occurred in concert with the exploitation of novel, UV-exposed habitats. Interestingly, these changes appear to have co-occurred with the transformation of the terrestrial landscape by gymnosperm- and angiosperm-dominated forests. Therefore, our results suggest the intriguing possibility that ecological opportunities related to global climate and vegetation also played a central role in the evolutionary origin of anthraquinones themselves.

Materials and Methods

Taxon and Locus Sampling. We selected taxa using our previous sampling of the Teloschistales (30) as a reference. Six loci were targeted for this study: nuclear ribosomal ITS, large subunit (nrLSU), and mitochondrial small subunit (mtSSU), and nuclear protein-coding genes *RPB1* (one locus) and *RPB2* (two amplicons, considered here as two separate loci). See [SI Appendix, Table S3](#) for voucher and GenBank accession number information, and see [SI Appendix, SI Materials and Methods](#) for details on alignments and datasets.

Phylogenetic Analyses. A synopsis of datasets used in our analyses is shown in [SI Appendix, Table S4](#). A total of 1,000 replicates of maximum-likelihood (ML) bootstrapping with a GTRGAMMA model were performed in RAXML (Version 7.2.8) (53, 54) to test for topological incongruence among each locus separately and on all possible combinations (55). For the *RPB2* locus, this criterion was applied on each amplicon separately. Phylogenetic relationships and confidence on the combined dataset were inferred primarily with Bayesian analyses by using MrBayes (Version 3.2.2) (56). In addition, support was also inferred using ML bootstrap proportions computed in RAXML (see [SI Appendix, SI Materials and Methods](#) for details on conflict and Bayesian and ML analyses settings).

Overall, a well-resolved and supported topology of the Teloschistales was recovered by both ML and Bayesian approaches on the six-locus combined dataset ([SI Appendix, Fig. S5](#)). The taxonomy of this group of lichens, specifically the family Teloschistaceae, has been highly controversial, and several classifications have recently been proposed with some newly delimited genera not significantly supported (e.g., refs. 29 and 57). On the grounds of practicality, here we use the traditional generic names for the group, recently included in a “without-prejudice” list of generic names of fungi for protection under the International Code of Nomenclature (58), being aware that their current circumscription is polyphyletic in some cases (see [SI Appendix, SI Materials and Methods](#) for more details).

Dating Analyses. The times of divergence were estimated by using a two-step approach. Because of the lack of fossils related to the group of interest, the results of a first dating analysis (eukaryotes dataset; [SI Appendix, Fig. S6](#)) was

used to obtain calibrations of the root and a few major clades using a subset of the Teloschistales. This first analysis was based on eight fossils, two secondary molecular-based calibrations across the eukaryotes, and several nodes constrained as monophyletic ([SI Appendix, Tables S5 and S6](#)). The second analysis then focused on the order Teloschistales by using time estimates from the first analysis as secondary calibrations to date divergences within the more exhaustive sampling of Teloschistales (Teloschistales dataset; [SI Appendix, Fig. S1 and Tables S7 and S8](#)). Settings for these two analyses are described in [SI Appendix, SI Materials and Methods](#). A relaxed-clock model of molecular evolution was allowed in both analyses.

Testing for Shifts in Diversification Rates. We used a Bayesian framework to model diversification using the BAMM program (59) and analyzed the results using the R package BAMMtools (36) to account for temporal and branch-specific shifts in diversification rates and to investigate whether regimes of net diversification (speciation rate – extinction rate) changed within the tree. Our dated phylogeny of the Teloschistales incorporates 108 species (Fig. 1A), ~10% of the currently ascribed total (29), but the polyphyletic nature of the very large *Caloplaca* genus means that it is impossible to reliably assign unsampled species to clades within the Teloschistaceae. In our analyses using BAMM, we therefore made estimates of our sampling frequency and accounted for missing species using clade-specific sampling fractions for our four major lineages as follows: 0.089 for Teloschistaceae, 0.116 for Megalosporaceae, 0.11 for Brigantiaaceae, and 0.176 for Letrouitiaceae. These fractions were based on current species estimates from the Index Fungorum database (accessed August 21, 2014) (see [SI Appendix, SI Materials and Methods](#) for details of the BAMM analyses).

Ancestral Character State Estimations. The traits anthraquinones in thallus, presence of anthraquinones, exposure to light, rock substrate, bark substrate, and growth form crustose-continuous were coded from our own sampled collections ([SI Appendix, Fig. S4 and Table S3](#)) and complemented with the literature. All traits were treated as binary and reconstructed on a selection of 40 nodes that represented interesting and well-supported clades by using, in addition to likelihood reconstruction, three separate methods that take into account possible biases caused by phylogenetic uncertainty, state-dependent differences in diversification rates and incomplete taxon sampling, and branch-dependent transition rates (using BayesTraits-Bayes, BiSSE-ML, and corHMM-ML, respectively), with the settings described in [SI Appendix, SI Materials and Methods](#). The plotting of different methods and traits can be found in [SI Appendix, Fig. S4](#), and model performance results are provided in [SI Appendix, Tables S1, S9, and S10](#). We established a statistical threshold to deal with discrepancies among methods and to determine for which nodes we could confidently estimate character states (Fig. 1A). For each trait and for each node, we calculated the mean estimated probability of trait presence across methods. When this value fell between 0.25 and 0.75, the state for that node was considered ambiguous and was excluded. We selected which methods to include in this calculation based on statistical tests of state- and branch-dependent diversification rates. Thus, for the crustose-continuous character, which showed a high significance for state-dependent diversification rates, we used only BiSSE results. Similarly, for shade, we used the two-rate model corHMM and BiSSE. For all other traits, we used all four methods.

Additionally, we performed a multistate reconstruction in BayesTraits treating the anthraquinones trait as polymorphic depending on the pigments' location. We wanted to test for directionality in the loss of anthraquinones, as initially hypothesized. The same settings as in the binary analyses were applied; several runs were performed, starting with RJ and trying different constraints and hyper priors. Here 15 models with different constraints describing trait evolution were compared that can be grouped in models allowing transitions and reversals with different rates; transitions and reversals with the same rate; and stepwise models where one-step changes have different rates than two-step changes, independently of the transitions and reversals. Also, the root was “fossilized” (i.e., fixed to each possible state) by using the ‘fossil’ command of BayesTraits. Log likelihoods and BFs were compared.

Testing for State-Dependent Diversification Rates. We also examined whether any of the reconstructed traits was correlated with increased rates of diversification using BiSSE (38). BiSSE uses ML to estimate absolute rates of asymmetric character change (q), speciation (λ), and/or extinction (μ) by maximizing the likelihood of these parameters for a given topology with branch lengths. For each character, we compared two models: (i) an unconstrained model in which q and λ were allowed to vary; and (ii) a constrained model in which λ and μ was set equal for each character. If diversification rates are

correlated with character states, then the unconstrained model should be favored over the constrained model. We carried out likelihood ratio tests (χ^2 approximation) and compared Δ AIC scores to assess the fit of models. Because the BiSSE model accounts for only the character change that occurs along lineages, we also implemented “BiSSE–node enhanced state shift” [BiSSE–ness (42)] model to allow for cladogenetic change in addition to anagenetic change, which thus enables us to address the mode of character change (along-lineage vs. speciation change) while at the same time accounting for the effect the character has on rates of speciation and extinction, i.e., trait-dependent diversification. We compared the fit of five different models, ranging from character change occurring only along lineages to when it occurred only at speciation, under different taxon sampling scenarios as described in *SI Appendix, SI Materials and Methods*.

- Glor RE (2010) Phylogenetic insights on adaptive radiation. *Annu Rev Ecol Syst* 41:251–270.
- Losos JB, Mahler DL (2010) Adaptive radiation: The interaction of ecological opportunity, adaptation, and speciation. *Evolution Since Darwin the First 150 Years*, eds Bell MA, Futuyma DJ, Eanes WF, Levinton JS (Sinauer, Sunderland, MA), pp 381–420.
- Losos JB (2010) Adaptive radiation, ecological opportunity, and evolutionary determinism. American Society of Naturalists E. O. Wilson award address. *Am Nat* 175(6): 623–639.
- Hughes C, Eastwood R (2006) Island radiation on a continental scale: Exceptional rates of plant diversification after uplift of the Andes. *Proc Natl Acad Sci USA* 103(27): 10334–10339.
- Schluter D (2000) *The Ecology of Adaptive Radiation* (Oxford Univ Press, Oxford).
- Hunter JP (1998) Key innovations and the ecology of macroevolution. *Trends Ecol Evol* 13(1):31–36.
- Heard SB, Hauser DL (1995) Key evolutionary innovations and their ecological mechanisms. *Hist Biol* 10(2):151–173.
- Kocher TD (2004) Adaptive evolution and explosive speciation: The cichlid fish model. *Nat Rev Genet* 5(4):288–298.
- Losos JB (2009) *Lizards in an Evolutionary Tree: Ecology and Adaptive Radiation of Anoles* (Univ of California Press, Oakland, CA).
- Ree RH (2005) Detecting the historical signature of key innovations using stochastic models of character evolution and cladogenesis. *Evolution* 59(2):257–265.
- Cracraft J (1990) The origin of evolutionary novelties: Pattern and process at different hierarchical levels. *Evolutionary Innovations*, ed Nitecki MH (Univ of Chicago Press, Chicago), pp 21–44.
- Hodges SA, Arnold ML (1995) Spurring plant diversification: Are floral nectar spurs a key innovation. *Proc Biol Sci* 262:343–348.
- Simpson GG (1953) *The Major Features of Evolution* (Columbia Univ Press, New York).
- Seehausen O (2007) Chance, historical contingency and ecological determinism jointly determine the rate of adaptive radiation. *Heredity (Edinb)* 99(4):361–363.
- Wagner CE, Harmon LJ, Seehausen O (2012) Ecological opportunity and sexual selection together predict adaptive radiation. *Nature* 487(7407):366–369.
- Lutzoni F, Miadlikowska J (2009) Lichens. *Curr Biol* 19(13):R502–R503.
- Nguyen K-H, Chollet-Krugler M, Gouault N, Tomasi S (2013) UV-protectant metabolites from lichens and their symbiotic partners. *Nat Prod Rep* 30(12):1490–1508.
- Hensen A, Jahns HM (1974) *Lichenes. Eine einföhrung in die Flechtenkunde* (Georg Thieme, Stuttgart).
- Elix JA, Stoker-Wörgötter E (2008) Biochemistry and secondary metabolites. *Lichen Biology*, ed Nash TH (Cambridge Univ Press, Cambridge, U.K.), 2nd Ed, pp 104–133.
- Rikkinen J (1995) *What's Behind the Pretty Colours?: A Study on the Photobiology of Lichens* (Bryobrothera 4, Helsinki).
- Boustie J, Tomasi S, Grube M (2011) Bioactive lichen metabolites: Alpine habitats as an untapped source. *Phytochem Rev* 10(3):287–307.
- Hauck M, Dulamsuren C, Mühlenberg M (2007) Lichen diversity on steppe slopes in the northern Mongolian mountain taiga and its dependence on microclimate. *Flora* 202:530–546.
- Bjerke JW, Lørfall K, Elvebakk A (2002) Effects of ultraviolet radiation and PAR on the content of usnic and divaricatic acids in two arctic-alpine lichens. *Photochem Photobiol Sci* 1(9):678–685.
- Rubio C, Fernández E, Hidalgo ME, Quilhot W (2002) Effects of solar UV-B radiation in the accumulation of rhizocarpic acid in a lichen species from Alpine zones of Chile. *Boletín de la Sociedad Chilena de Química* 47(1):67–72.
- Solhaug KA, Gauslaa Y, Nybakken L, Bilger W (2003) UV-induction of sun-screening pigments in lichens. *New Phytol* 158(1):91–100.
- Solhaug KA, Gauslaa Y (2004) Photosynthetic stimulation of the UV-B induced fungal anthraquinone synthesis in the foliose lichen *Xanthoria parietina*. *Plant Cell Environ* 27(2):167–176.
- Brunauer G, Muggia L, Stoker-Wörgötter E, Grube M (2009) A transcribed polyketide synthase gene from *Xanthoria elegans*. *Mycol Res* 113(Pt 1):82–92.
- Kirk PM, Cannon PF, Minter DW, Stalpers JA (2008) *Dictionary of the Fungi* (CABI, Wallingford, U.K.), 10th Ed.
- Arup U, Sochting U, Frøden P (2013) A new taxonomy of the family Teloschistaceae. *Nordic J Bot* 31(1):016–083.
- Gaya E, et al. (2012) Implementing a cumulative supermatrix approach for a comprehensive phylogenetic study of the Teloschistales (Pezizomycotina, Ascomycota). *Mol Phylogenet Evol* 63(2):374–387.
- Behrensmeyer AK, et al. (1992) *Terrestrial Ecosystems Through Time: Evolutionary Paleocology of Terrestrial Plants and Animals*, eds Behrensmeyer AK, et al. (Univ of Chicago Press, Chicago).
- Amo de Paz G, Cubas P, Divakar PK, Lumbsch HT, Crespo A (2011) Origin and diversification of major clades in parmelioid lichens (Parmeliaceae, Ascomycota) during the Paleogene inferred by Bayesian analysis. *PLoS One* 6(12):e28161.
- Miadlikowska J, et al. (2014) A multigenic phylogenetic synthesis for the class Lecanoromycetes (Ascomycota): 1307 fungi representing 1139 infrageneric taxa, 317 genera and 66 families. *Mol Phylogenet Evol* 79:132–168.
- Alley NF, Frakes LA (2003) First known Cretaceous glaciation: Livingston Tillite member of the Cadna-owie Formation, South Australia. *Aust J Earth Sci* 50(2):139–144.
- Chumakov NM (2004) Trends in global climate changes inferred from geological data. *Stratigr Geol Correl* 12(2):117–138.
- Rabosky DL, Donnellan SC, Grundler M, Lovette IJ (2014) Analysis and visualization of complex macroevolutionary dynamics: An example from Australian scincid lizards. *Syst Biol* 63(4):610–627.
- Pagel M, Meade A, Barker D (2004) Bayesian estimation of ancestral character states on phylogenies. *Syst Biol* 53(5):673–684.
- Maddison WP, Midford PE, Otto SP (2007) Estimating a binary character's effect on speciation and extinction. *Syst Biol* 56(5):701–710.
- Beaulieu JM, Donoghue MJ (2013) Fruit evolution and diversification in campanulid angiosperms. *Evolution* 67(11):3132–3144.
- Beaulieu JM, O'Meara BC, Donoghue MJ (2013) Identifying hidden rate changes in the evolution of a binary morphological character: The evolution of plant habit in campanulid angiosperms. *Syst Biol* 62(5):725–737.
- FitzJohn RG, Maddison WP, Otto SP (2009) Estimating trait-dependent speciation and extinction rates from incompletely resolved phylogenies. *Syst Biol* 58(6):595–611.
- Magnusson-Ford K, Otto SP (2012) Linking the investigations of character evolution and species diversification. *Am Nat* 180(2):225–245.
- Graham A (2011) The age and diversification of terrestrial New World ecosystems through Cretaceous and Cenozoic time. *Am J Bot* 98(3):336–351.
- McLoughlin S (2001) The breakup history of Gondwana and its impact on pre-Cenozoic floristic provincialism. *Aust J Bot* 49(3):271–300.
- Magallón S, Hilu KW, Quandt D (2013) Land plant evolutionary timeline: Gene effects are secondary to fossil constraints in relaxed clock estimation of age and substitution rates. *Am J Bot* 100(3):556–573.
- Smith SA, Beaulieu JM, Donoghue MJ (2010) An uncorrelated relaxed-clock analysis suggests an earlier origin for flowering plants. *Proc Natl Acad Sci USA* 107(13):5897–5902.
- Bell CD, Soltis DE, Soltis PS (2010) The age and diversification of the angiosperms revisited. *Am J Bot* 97(8):1296–1303.
- Clarke JT, Warnock RCM, Donoghue PCJ (2011) Establishing a time-scale for plant evolution. *New Phytol* 192(1):266–301.
- Schneider H, et al. (2004) Ferns diversified in the shadow of angiosperms. *Nature* 428(6982):553–557.
- Schuettpelz E, Pryer KM (2009) Evidence for a Cenozoic radiation of ferns in an angiosperm-dominated canopy. *Proc Natl Acad Sci USA* 106(27):11200–11205.
- Wikström N, Kenrick P (2001) Evolution of Lycopodiaceae (Lycopodiaceae): Estimating divergence times from *rbcL* gene sequences by use of nonparametric rate smoothing. *Mol Phylogenet Evol* 19(2):177–186.
- Prieto M, Wedin M (2013) Dating the diversification of the major lineages of Ascomycota (Fungi). *PLoS One* 8(6):e65576.
- Stamatakis A (2006) RAxML-VI-HP: Maximum likelihood-based phylogenetic analyses with thousands of taxa and mixed models. *Bioinformatics* 22(21):2688–2690.
- Stamatakis A, Hoover P, Rougemont J (2008) A rapid bootstrap algorithm for the RAxML Web servers. *Syst Biol* 57(5):758–771.
- Mason-Gamer RJ, Kellogg EA (1996) Testing for phylogenetic conflict among molecular data sets in the tribe Triticeae (Gramineae). *Syst Biol* 45(4):524–545.
- Ronquist F, et al. (2012) MrBayes 3.2: Efficient Bayesian phylogenetic inference and model choice across a large model space. *Syst Biol* 61(3):539–542.
- Kondratyuk SY, Kärnefelt I, Thell A, Elix JA (2014) A revised taxonomy for the subfamily Caloplacoidae (Teloschistaceae, Ascomycota) based on molecular phylogeny. *Acta Bot Hung* 56(1–2):93–123.
- Kirk PM, et al. (2013) A without-prejudice list of generic names of fungi for protection under the International Code of Nomenclature for algae, fungi, and plants. *IMA Fungus* 4(2):381–443.
- Rabosky DL (2014) Automatic detection of key innovations, rate shifts, and diversity-dependence on phylogenetic trees. *PLoS One* 9(2):e89543.