

New Phytologist Supporting Information Fig. S1, Tables S1–S3 and Methods S1

Article title: Biological nitrogen fixation by alternative nitrogenases in boreal cyanolichens: importance of molybdenum availability and implications for current biological nitrogen fixation estimates

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Fig. S1 Unwashed thallus and cephalodia content for V and Mo.

Table S1 Discrimination of lichen compartment with respect to metals

Table S2 Regression parameters for linear regression in Figs 2, 3(e) and 5(b)

Table S3 Literature survey of BNF estimation and measurement conditions for di-nitrogen fixing species in various ecosystems around the world

Methods S1 Contribution of alternative nitrogenase to acetylene reduction and to N₂ fixation.

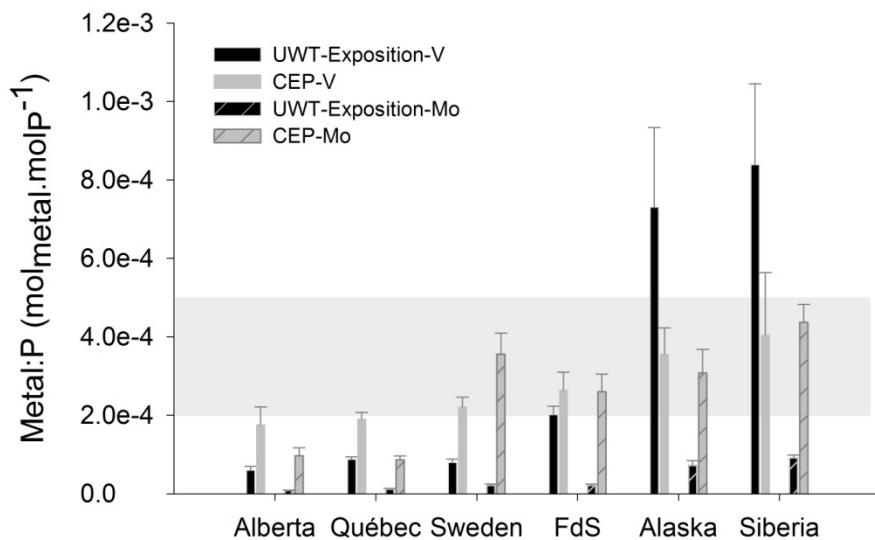
Methods S1 Contribution of alternative nitrogenase to acetylene reduction ($f_{\text{alt ara}}$) and to N₂ fixation ($f_{\text{alt N}_2}$).

$$f_{\text{alt ara}} = \frac{(^{13}\varepsilon_{\text{Mo}} - ^{13}\varepsilon_{\text{AR, sample}})}{(^{13}\varepsilon_{\text{Mo}} - ^{13}\varepsilon_{\text{alt}})}$$

$$f_{\text{alt N}_2} = \frac{(f_{\text{alt ara}}/1)}{\left(\frac{f_{\text{alt ara}}}{1}\right) + \left(\frac{1 - f_{\text{alt ara}}}{3}\right)}$$

Contributions were calculated according to Zhang *et al.* (2016): Isozyme specific $^{13}\varepsilon$ (see main text) for Mo and V were determined in deletion mutants CA 1.70 (Mo-Nase only) and CA 11.70 (V-Nase only) of *Azotobacter vinelandii*, in *Rhodopseudomonas palustris* wild type (CGA009) and deletion mutant CGA753 (Mo-Nase only) and CGA766 (V-Nase only), as well as in a wild type strain of *Anabaena variabilis* (ATCC 29413) as $^{13}\varepsilon_{\text{Mo}} = 13.8 \pm 0.3\%$ and $^{13}\varepsilon_{\text{V}} = 7.9 \pm 0.2\%$. Value of $^{13}\varepsilon_{\text{alt}}$ for V was used for all calculations of $f_{\text{alt ara}}$ in this manuscript.

a



b

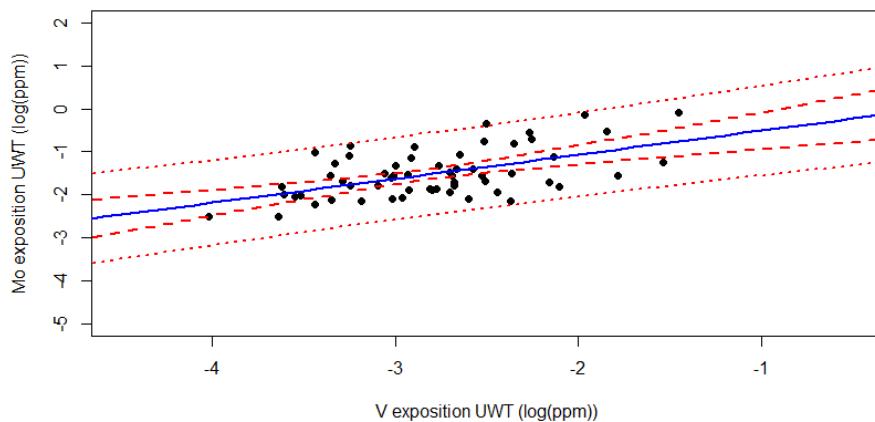


Fig. S1 Unwashed thallus (UWT, i.e. metal exposure) and cephalodia (CEP) content for V and Mo. (a) Metal content of UWT and CEP on sampling sites expressed in mol_V.mol_P⁻¹ (modification of Darnajoux *et al.*, 2014) pale background represents optimal V and Mo cellular quota required to sustain diazotrophic growth (more details in Darnajoux *et al.*, 2014). FdS stands for Fjord-du-Saguenay, South Québec. (b) Linear regression (solid line) of Mo exposition ($\mu\text{g}_{\text{Mo}} \cdot \text{g}_{\text{thallus}}^{-1}$) and V exposition ($\mu\text{g}_{\text{Mo}} \cdot \text{g}_{\text{thallus}}^{-1}$) in lichen thallus in nonheavily contaminated boreal sites (Alberta, Québec, FdS and Sweden). Short dash lines show 95% prediction intervals and long dash lines show 95% confidence intervals. All data were log-transformed to achieve normality of residues and to limit spurious correlation.

Table S1 Discrimination of lichen compartment with respect to metals

Values prediction	n	Recall	Precision	
WT	50	0.9400		0.0408
UWT	49	0.9592		0.0600
CEP	49	0.9796		0.0000
ALG	47	1.0000		0.0208
Error rate			0.0308	
Classification functions	WT	UWT	CEP	ALG
Mg	15.16	25.48	7.01	30.10
Ti	1.20	0.12	-5.41	1.24
V	-48.44	-50.95	-43.32	-49.96
Cr	-0.01	-1.80	2.39	1.26
Mn	6.20	10.34	4.91	4.90
Fe	43.38	41.72	45.95	41.08
Co	2.53	3.05	-0.94	2.16
Ni	-16.64	-15.92	-9.65	-15.73
Zn	-1.44	-4.00	-1.24	-5.36
Mo	-14.65	-16.11	-8.61	-11.36
Cd	-29.02	-28.81	-26.77	-29.45
Pb	-5.61	-2.30	-9.45	-4.91
constant	-470.15	-476.36	-396.87	-454.57

Confusion matrix (leave-one-out method) and classification function from linear discriminant analysis of thallus compartment of cyanolichen *P.aphthosa* s.l. (UWT, unwashed thallus (i.e. exposition); WT, oxalate-EDTA washed thallus; ALG, algae; CEP, Cephalodia). Data in metal : P ratio were log-transform to avoid spurious correlation and achieved normality.

Table S2 Regression parameters for linear regression in Figs 2, 3(e) and 5(b)

Regression	Model equation	n	P-value
<u>Fig. 2 (Sample from Alberta, South Québec (FdS), North Québec and Sweden)</u>			
$\log_{10}(Mo:P (mol_{Mo} \cdot mol_P^{-1})) = f(\log_{10}(Mo \text{ UWT} (\mu g_{Mo} \cdot g_{thallus}^{-1})))$	0.79 ± 0.14 × log₁₀(Mo UWT) - 2.94 ± 0.16	58	<0.001
$\log_{10}(V:P (mol_V \cdot mol_P^{-1})) = f(\log_{10}(V \text{ UWT} (\mu g_V \cdot g_{thallus}^{-1})))$	0.06 ± 0.10 × log₁₀(V UWT) - 3.71 ± 0.07	58	0.563
$\log_{10}(V:P (mol_V \cdot mol_P^{-1})) = f(\log_{10}(Mo:P (mol_{Mo} \cdot mol_P^{-1})))$	-0.61 ± 0.17 × log₁₀((Mo : P)²) - 4.7 ± 1.3 × log₁₀(Mo : P) - 13 ± 3	58	0.003
<u>Fig. 3e</u>			
$\log_{10}(Mo:P \text{ UWT} (mol_{Mo} \cdot mol_P^{-1})) = f(\log_{10}(V:P \text{ UWT} (mol_V \cdot mol_P^{-1})))$	0.69 ± 0.09 × log₁₀(V:P) - 0.9 ± 0.4	16	<0.001
$\log_{10}(Mo:P \text{ CEP} (mol_{Mo} \cdot mol_P^{-1})) = f(\log_{10}(V:P \text{ CEP} (mol_V \cdot mol_P^{-1})))$	-0.61 ± 0.16 × log₁₀(V:P) - 5.9 ± 0.6	16	0.002
<u>Fig. 5b (Grouped per Time since last fire)</u>			
$\delta 15N \text{ WT} = f(V:P (mol_V \cdot mol_P^{-1}) \text{ CEP})$	-0.00009 ± 0.00002 × (V:P) + 0.00027 ± 0.00002	14	0.023

*Bold figures highlight important parameter of the regression. Uncertainties are ± SE for the regression parameters.

Table S3 Literature survey of BNF estimates and measurement conditions for di-nitrogen fixing species in various biomes around the world; studies were screened from two review articles on biological nitrogen fixation (Cleveland *et al.*, 1999; Elbert *et al.*, 2012), with emphasis on mosses and lichens from boreal and arctic area

Geography	Details	Species	BNF estimation (kgN ha ⁻¹ yr ⁻¹)	ARA	ARA condition	¹⁵ N calibration	¹⁵ N condition	C ₂ H ₂ : N ₂ (mol mol ⁻¹)	References
New Mexico	Alpine climate, Douglas fir, spruce fir	<i>Peltigera</i> spp.	0.04-3.3	No	---	No	---	Forman 1975 estimations	Forman & Dowden (1977)
West North Carolina	---	<i>Lobaria pulmonaria</i> , <i>Lobaria quercizans</i>	0.8	Yes	Field (1 h)	No	---	N.C.	Becker (1980)
New Zealand	Urewera National Park,	<i>Sticta</i> , <i>Pseudocyphellaria</i>	1 - 10	Yes	26°C, 1 h 30–3 h	Yes	26°C, 1 h 30–3 h	C ₂ H ₂ = 5.6 × N ₂ -6.3	Green <i>et al.</i> (1980)
Scotland and South Wales	---	<i>Peltigera membranacea</i> , <i>P. polydactyla</i> , <i>Lobaria pulmonaria</i>	2.4-5.8	Yes	Climatic chamber 2.5–18°C	Yes	Climatic chamber 2.5–18°C	7.8-12.2	Millbank (1981)
Nebraska	Prairies	Cyanolichens, heterotrophs	3.5	Yes	18–22°C	No	---	3	Kapustka & DuBois (1987)
Northwest Pacific	Canopy	<i>Lobaria oregana</i>	1.5-16.5	Yes	Field/Lab (0–20°C)	No	---	4	Antoine (2004)
Puerto Rico	Tropical forest	Soil, phyllophyte, moss, lichens	8.4-12.3	Yes	Field	No	---	3	Cusack <i>et al.</i> (2009)
South-central Chile	Rain forest chronosequence	Soil, cyanolichens, moss	0-6	Yes	Lab	No	---	3	Pérez <i>et al.</i> (2014)
Sweden, Finland, Norway	Boreal forest	<i>Pleurozium schreberi</i>	1.5-2	Yes	Field	Yes	Lab	3	DeLuca <i>et al.</i> (2002)
Northern Sweden	Boreal forest chronosequence	<i>Pleurozium schreberi</i> , <i>Hylocomnium splendens</i>	1.6	Yes	---	ND	---	3	Zackrisson <i>et al.</i> (2009)
Northern Sweden	Boreal forest chronosequence	<i>Pleurozium schreberi</i>	0.2-7	Yes	---	No	---	3	DeLuca <i>et al.</i> (2007)
Northern Sweden	Boreal forest chronosequence	<i>Pleurozium schreberi</i>	0.5-2	Yes	---	Yes	Lab	3	Zackrisson <i>et al.</i> (2004)

Northern Sweden	Boreal forest	<i>Pleurozium schreberi</i> , <i>Hylocomnium splendens</i>	0.52-2	Yes	---	Yes (<i>H.splendens</i>)	Lab	3	Lagerström <i>et al.</i> (2007)
Northern Sweden	Sub-arctic	Moss, Lichens	0.6-2.6	No	---	Yes	Field 7–23°C	---	Gavazov <i>et al.</i> (2010)
Northern Sweden	Sub-arctic	<i>Hylocomnium splendens</i> <i>Peltigera aphthosa</i> <i>Sphagnum</i>	0.3 0.9 2.6	Yes	Field (10–35°C)	No	---	2.78	Rousk <i>et al.</i> (2015)
Northern Sweden	Sub-arctic	<i>Sphagnum, Drepanocladus</i>	94	Yes	---	No	---	3	Granhall & Selander (1973)
Northern Sweden	Sub-arctic	<i>Sphagnum</i>	32	Yes	---	No	---	3	Basilier & Granhall (1978)
British Colombia	Rain forest	Moss	0.26-0.76	Yes	16°C	No	---	3	Lindo & Whiteley (2011)
Arctic	High Arctic	Soil and cyanobacterial mat	0.6-1	Yes	Field	No	---	3	Henry & Svoboda (1986)
Ohio	Prairies	<i>Nostoc</i> sp.	4.6+3.19	Yes	Field (9–30°C)	No	---	3	Dubois & Kapustka (1983)
Inner Mongolia	Steppe	<i>Nostoc</i> sp., lichens	0.033-0.087	Yes	25°C	Yes	5°C and 25°C	0.31	Holst <i>et al.</i> (2009)
Southern Utah	---	Biocrust	0.02-3.63	Yes	21–22.5°C	No	---	3	Jeffries <i>et al.</i> (1992)
Indiana, South Lake Michigan	Sand dune soil	Soilcrust	0.2-8	No	---	No	Mass balance	---	Thiet <i>et al.</i> (2005)
China, Loess plateau	Soil, grassland	Biocrust	4-13	Yes	Growth chambers (5–45°C)	No	---	3	Zhao <i>et al.</i> (2010)
Southern Utah	---	Biocrust	1.4-13	Yes	26°C	No	---	0.062	Belnap (2002)
Southern Alberta	---	<i>Nostoc</i> sp.	0.1-0.773 (31days)	Yes	Field with water bath	No	---	3	Coxson & Kershaw (1983)
Niger	---	Biocrust	3.5	Yes	30°C	No	---	NC	Malam Issa <i>et al.</i> (2001)
Northwest China	Desert	Biological soil crust	C2H2 data only	Yes	26°C (24 h)	No	---	3	Wu <i>et al.</i> (2009)

Zambia, Botswana	---	Soil crust	0.008-0.044	Yes	Field (48 h)	No	---	3	Aranibar <i>et al.</i> (2003)
Costa Rica	Rain forest	<i>Scytonema</i> sp.	2-5	Yes	Field	No	---	4	Freiberg (1998)
French Guiana	Tropical forest	Cyanobacteria biofilm	134-233	No	---	No	Value from Freiberg 1998	---	Dojani <i>et al.</i> (2007)
North western Ohio	---	Biocrust	1.3	Yes	26°C (4 h)	No	---	3.2	Veluci <i>et al.</i> (2006)
Western North Carolina	---	Woody litter, soil, phyllosphere, leaf litter	12.04	Yes	18°C (16–24 h)	No	---	NC	Todd <i>et al.</i> (1978)
Papua New Guinea, Australia	---	Phyllosphere	0.5	Yes	Field	No	---	3	Goosem & Lamb (1986)
California	---	Soil	2.1-4.8	Yes	Lab (26°C, 1 h)	Yes	Lab (26°C, 1 h)	3.1-8.4	Steyn & Delwiche (1970)
Central Sweden	Boreal forest	Soil	0.4-1.4	Yes	Lab (20°C)	Yes	Lab (20°C)	1.6-5.6	Nohrstedt (1985)
Ontario	Boreal transition zone	Soil	0.02-0.26	Yes	30°C	No	---	3	Hendrickson (1990)
Hawaii	---	Soil	0.06-1.29	Yes	Lab	Yes	---	3.9	Vitousek & Hobbie (2000)
Oregon	---	Wood log	1.4	Yes	Lab 22°C	Yes	Lab 22°C	3.5	Silvester <i>et al.</i> (1982)
Oregon	---	Wood roots	6.3	Yes	Lab 22°C	Yes	Lab 22°C	3.5	Chen & Hicks (2003)
Hawaii	---	Leaf litter, roots, soil	0.1-4.9	Yes	Lab (12–28°C)	Yes	Lab, 25°C	1.07-12.1	Ley & D'Antonio (1998)
Germany (Bavaria)	---	soil	0.2	Yes	Lab (20°C)	Yes	Lab	6-8	Limmer & Drake (1996)
Oregon	Andrew experimental forest	<i>Lobaria oregana</i>	3.5	Yes	Field (1 h)	No	---	3	Denison (1979)

Hawaii	Kilauea volcano	<i>Stereocaulon volcani</i> , leaf litter, liverworts	0.3-2.8	Yes	Field (24h)	Yes	Lab?	3.1/ 3.8 / 5.4	Vitousek (1994)
New Zealand	60000 yr chronosequence	<i>Coriaria</i> / lichens/moss/litter	11/0.02-2/0.7-9.6/1.1-1.9	Yes	Field	Yes	Field	1.33/1.58/0. 25	Menge & Hedin (2009)

References

- Antoine ME.** 2004. An ecophysiological approach to quantifying nitrogen fixation by *Lobaria oregana*. *The Bryologist* **107**: 82–87.
- Aranibar J, Anderson I, Ringrose S, Macko S.** 2003. Importance of nitrogen fixation in soil crusts of southern African arid ecosystems: acetylene reduction and stable isotope studies. *Journal of Arid Environments* **54**: 345–358.
- Basilier K, Granhall U.** 1978. Nitrogen fixation in wet minerotrophic moss communities of a subarctic mire. *Oikos* **31**: 236–246.
- Becker VE.** 1980. Nitrogen fixing lichens in forests of the southern appalachian mountains of North Carolina. *Bryologist* **83**: 29–39.
- Belnap J.** 2002. Nitrogen fixation in biological soil crusts from southeast Utah, USA. *Biology and Fertility of Soils* **35**: 128–135.
- Chen H, Hicks W.** 2003. High asymbiotic N₂ fixation rates in woody roots after six years of decomposition: controls and implications. *Basic and Applied Ecology* **4**: 479–486.
- Cleveland CC, Townsend AR, Schimel DS, Fisher H, Hedin LO, Perakis S, Latty EF, Von Fischer C, Elseroad A, Wasson MF.** 1999. Global patterns of terrestrial biological nitrogen (N₂) fixation in natural ecosystems. *Global Biochemical Cycles* **13**: 623–645.
- Coxson DS, Kershaw KA.** 1983. The pattern of *in situ* summer nitrogenase activity in terrestrial *Nostoc* commune from *Stipa–Bouteloua* grassland, southern Alberta. *Canadian Journal of Botany* **61**: 2686–2693.
- Cusack DF, Silver W, McDowell WH.** 2009. Biological nitrogen fixation in two tropical forests: ecosystem-level patterns and effects of nitrogen fertilization. *Ecosystems* **12**: 1299–1315.
- Darnajoux R, Constantin J, Miadlikowska J, Lutzoni F, Bellenger J-P.** 2014. Is vanadium a biometal for boreal cyanolichens? *New Phytologist* **202**: 765–771.
- DeLuca TH, Zackrisson O, Gentili F, Sellstedt A, Nilsson MC.** 2007. Ecosystem controls on nitrogen fixation in boreal feather moss communities. *Oecologia* **152**: 121–130.
- DeLuca TH, Zackrisson O, Nilsson M-C, Sellstedt A.** 2002. Quantifying nitrogen-fixation in feather moss carpets of boreal forest. *Nature* **419**: 917–920.
- Denison WC.** 1979. *Lobaria oregana*, a nitrogen-fixing lichen in old-growth Douglas fir forests. In: Gordon JC, Wheeler CT, Perry DA, eds. *Symbiotic nitrogen fixation in the management of temperate forests*. Corvallis, OR, USA: Forest Research Laboratory, Oregon State University, 266–275.

Dojani S, Lakatos M, Rascher U, Wanek W, Lüttge U, Büdel B. 2007. Nitrogen input by cyanobacterial biofilms of an inselberg into a tropical rainforest in French Guiana. *Flora* **202**: 521–529.

Dubois J, Kapustka L a. 1983. Biological nitrogen influx in an Ohio relict prairie. *American Journal of Botany* **70**: 8–16.

Elbert W, Weber B, Burrows S, Steinkamp J, Büdel B, Andreae MO, Pöschl U. 2012. Contribution of cryptogamic covers to the global cycles of carbon and nitrogen. *Nature Geoscience* **5**: 459–462.

Forman RTT, Dowden DL. 1977. Nitrogen fixing lichen roles from desert to alpine in the Sangre de Cristo mountains, New Mexico. *The Bryologist* **80**: 561–570.

Freiberg E. 1998. Microclimatic parameters influencing nitrogen fixation in the phyllosphere in a Costa Rican premontane rain forest. *Oecologia* **117**: 9–18.

Gavazov KS, Soudzilovskaia NA, van Logtestijn RSP, Braster M, Cornelissen JHC. 2010. Isotopic analysis of cyanobacterial nitrogen fixation associated with subarctic lichen and bryophyte species. *Plant and Soil* **333**: 507–517.

Goosem S, Lamb D. 1986. Measurements of phyllosphere nitrogen fixation in a tropical and two sub-tropical rain forests. *Journal of Tropical Ecology* **2**: 373.

Granhall U, Selander H. 1973. Nitrogen fixation in a subarctic mire. *Oikos* **24**: 8–15.

Green TGA, Horstmann J, Bonnett H, Wilkins A, Silvester WB. 1980. Nitrogen fixation by members of the *Stictaceae* (Lichenes) of New Zealand. *New Phytologist* **84**: 339–348.

Hendrickson OQ. 1990. Asymbiotic nitrogen fixation and soil metabolism in three Ontario forests. *Soil Biology and Biochemistry* **22**: 967–971.

Henry GHR, Svoboda J. 1986. Dinitrogen fixation (acetylene reduction) in high Arctic sedge meadow communities. *Arctic & Alpine Research* **18**: 181–187.

Holst J, Butterbach-Bahl K, Liu C, Zheng X, Kaiser AJ, Schnitzler JP, Zechmeister-Boltenstern S, Brüggemann N. 2009. Dinitrogen fixation by biological soil crusts in an Inner Mongolian steppe. *Biology and Fertility of Soils* **45**: 679–690.

Jeffries DL, Klopatek JM, Link SO, Bolton H. 1992. Acetylene reduction by cryptogamic crusts from a blackbrush community as related to resaturation and dehydration. *Soil Biology and Biochemistry* **24**: 1101–1105.

Kapustka LA, DuBois JD. 1987. Dinitrogen fixation by cyanobacteria and associative rhizosphere bacteria in the Arapaho Prairie in the Sand Hills of Nebraska. *American Journal of Botany* **74**: 107.

Lagerström A, Nilsson MC, Zackrisson O, Wardle DA. 2007. Ecosystem input of nitrogen through biological fixation in feather mosses during ecosystem retrogression. *Functional Ecology* **21**: 1027–1033.

Ley RE, D'Antonio CM. 1998. Exotic grass invasion alters potential rates of N fixation in Hawaiian woodlands. *Oecologia* **113**: 179–187.

Limmer C, Drake HL. 1996. Non-symbiotic N₂-fixation in acidic and pH-neutral forest soils: aerobic and anaerobic differentials. *Soil Biology and Biochemistry* **28**: 177–183.

Lindo Z, Whiteley JA. 2011. Old trees contribute bio-available nitrogen through canopy bryophytes. *Plant and Soil* **342**: 141–148.

Malam Issa O, Stal LJ, Défarge C, Couté A, Trichet J. 2001. Nitrogen fixation by microbial crusts from desiccated Sahelian soils (Niger). *Soil Biology and Biochemistry* **33**: 1425–1428.

Menge DNL, Hedin LO. 2009. Nitrogen fixation in different biogeochemical niches along a 120 000-year chronosequence in New Zealand. *Ecology* **90**: 2190–2201.

Millbank JW. 1981. The assessment of nitrogen fixation and throughput by lichens. I The use of a controlled environment chamber to relate acetylene reduction estimates to nitrogen fixation. *New Phytologist* **89**: 647–655.

Nohrstedt H-Ö. 1985. Nonsymbiotic nitrogen fixation in the topsoil of some forest stands in central Sweden. *Canadian Journal of Forest Research* **15**: 715–722.

Pérez CA, Thomas FM, Silva WA, Segura B, Gallardo B, Armesto JJ. 2014. Patterns of biological nitrogen fixation during 60 000 years of forest development on volcanic soils from south-central Chile. *New Zealand Journal of Ecology* **38**: 189–200.

Rousk K, Sorensen PL, Lett S, Michelsen A. 2015. Across-habitat comparison of diazotroph activity in the subarctic. *Microbial Ecology* **69**: 778–787.

Silvester WB, Sollins P, Verhoeven T, Cline SP. 1982. Nitrogen fixation and acetylene reduction in decaying conifer boles: effects of incubation time, aeration, and moisture content. *Canadian Journal of Forest Research* **12**: 646–652.

Steyn PL, Delwiche CC. 1970. Nitrogen fixation by nonsymbiotic microorganisms in some California soils. *Environmental Science & Technology* **4**: 1122–1128.

Thiet RK, Boerner REJ, Nagy M, Jardine R. 2005. The effect of biological soil crusts on throughput of rainwater and N into Lake Michigan sand dune soils. *Plant and Soil* **278**: 235–251.

Todd RL, Meyer RD, Waide JB. 1978. Nitrogen fixation in a deciduous forest in the southeastern United States. *Ecological Bulletins* **26**: 172–177.

- Veluci RM, Neher DA, Weicht TR. 2006.** Nitrogen fixation and leaching of biological soil crust communities in mesic temperate soils. *Microbial Ecology* **51**: 189–196.
- Vitousek PM. 1994.** Potential nitrogen fixation during primary succession in Hawaii volcanoes national park. *Biotropica* **26**: 234–240.
- Vitousek PM, Hobbie S. 2000.** Heterothrophic nitrogen fixation in decomposing litter: patterns and regulation. *Ecology* **81**: 2366–2376.
- Wu N, Zhang YM, Downing a. 2009.** Comparative study of nitrogenase activity in different types of biological soil crusts in the Gurbantunggut Desert, Northwestern China. *Journal of Arid Environments* **73**: 828–833.
- Zackrisson O, DeLuca TH, Gentili F, Sellstedt A, Jäderlund A. 2009.** Nitrogen fixation in mixed *Hylocomnium splendens* moss communities. *Oecologia* **161**: 15–24.
- Zackrisson O, DeLuca TH, Nilsson M-C, Sellstedt A, Berglund LM. 2004.** Nitrogen fixation increases with successional age in boreal forests. *Ecology* **85**: 3327–3334.
- Zhang X, McRose DL, Darnajoux R, Bellenger J-P, Morel FMM, Kraepiel AML. 2016.** Alternative nitrogenase activity in the environment and nitrogen cycle implications. *Biogeochemistry* **127**: 189–198.
- Zhao Y, Xu M, Belnap J. 2010.** Potential nitrogen fixation activity of different aged biological soil crusts from rehabilitated grasslands of the hilly Loess Plateau, China. *Journal of Arid Environments* **74**: 1186–1191.